

UC Davis Office of the Attending Veterinarian Standards of Care

Policy: SC-40-102
Date: 6/7/2016
Enabled by: The Guide,
APHIS, AVMA,
IACUC /AV
Supersedes: IACUC Policy,
ALL Previous
Standards of Care
on Euthanasia

Title: Euthanasia Procedures for the UC Davis Animal Care Program

I. Purpose:

The purpose of this policy is to establish minimum standards for euthanasia for species in the lab animal setting.

II. Policy:

All units providing animal care for rodent species must meet or exceed these minimum requirements for euthanasia based on the *Guide for the Care and Use of Laboratory Animals*, the *Ag Guide* and the *AVMA Guidelines for the Euthanasia of Animals*. Research staff shall use the methods approved in their animal protocol. Veterinary Staff and Husbandry staff may use an AVMA approved method as described below, provided the appropriate training and certification have been completed.

III. Procedure:

Refer to the AVMA Guidelines on Euthanasia for approved euthanasia methods:

<http://www.avma.org/KB/policies/Documents/euthanasia.pdf>

The agents and methods of euthanasia appropriate for rodent species are available in the AVMA Guidelines for the Euthanasia of Animals: 2013 Edition or subsequent revisions of that document. Euthanasia is the procedure of killing an animal rapidly, painlessly, and without distress.

Euthanasia must be carried out by trained personnel using acceptable techniques in accordance with applicable regulations and policies. The method used should not interfere with postmortem evaluations. Proper euthanasia involves skilled personnel to help ensure that the technique is performed humanely and effectively and to minimize risk of injury to people. Personnel who perform euthanasia must have training and experience with the techniques to be used. The equipment and materials required to

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perform euthanasia should be readily available, and the attending veterinarian or a qualified animal scientist/trainer should ensure that all personnel performing euthanasia have demonstrated proficiency in the use of the techniques selected.

Euthanasia techniques should result in rapid unconsciousness followed by cardiac or respiratory arrest and the ultimate loss of brain function. In addition, the technique used should minimize any stress and anxiety experienced by the animal before unconsciousness. For this reason, anesthetic agents are generally acceptable, and animals of most species can be quickly and humanely euthanized with the appropriate injection of an overdose of a barbiturate. Certain other methods may be used for euthanasia of anesthetized animals because the major criterion (insensibility) has been fulfilled.

Physical methods of euthanasia may be used if persons performing the procedure are properly trained. If physical methods such as cervical dislocation are used, anesthetizing the rodent prior to this procedure is recommended. Every attempt should be made to minimize stress to the animal before euthanasia.

If euthanizing more than one animal at a time in the same chamber, the chamber must not be overcrowded. Animals must be able to move freely and make normal postural adjustments. Only animals of the same species may be euthanized in the same chamber at the same time.

Regardless of the method of euthanasia that is performed, personnel must ensure that death has occurred. A combination of criteria is most reliable in confirming death, including lack of pulse, breathing, corneal reflex and response to response to firm toe pinch, inability to hear respiratory sounds and heartbeat by use of a stethoscope, graying of the mucous membranes and rigor mortis. None of these signs alone, except rigor mortis, confirms death.

Personnel shall be trained on how to assure death in animals.

2013 AVMA Guidelines for the Euthanasia of Animals

Rodent Species:

Acceptable Methods	Acceptable with Conditions
Injected barbiturates and barbiturate combinations Injected dissociative agent combinations	Inhaled anesthetics CO ₂ ** CO Tribromoethanol Cervical dislocation Decapitation Focused beam microwave irradiation

**Conditions required for CO₂ euthanasia provided

-Source is from a compressed gas CO₂ cylinder (dry ice and other sources not acceptable)

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- Flow rate displaces 10%-30% of the chamber volume per minute (prefilled chambers are not acceptable)
- Flow should be maintained for one minute after respiratory arrest
- Death is verified by physical exam or ensured by an adjunctive physical method
- Animals should be euthanized in their home cages whenever possible
- Chamber should be cleaned after each use

Dogs and Cats:

Acceptable	Acceptable With Conditions
Intravenous barbiturates, injected anesthetic overdose	Barbiturates (alternate routes of administration), inhaled anesthetic overdose

Rabbits:

Acceptable	Acceptable With Conditions
Intravenous barbiturates	Inhaled anesthetic overdose, CO ₂ , cervical dislocation (as anatomically appropriate), penetrating captive bolt

Finfish:

Acceptable	Acceptable with Conditions
Immersion in buffered benzocaine or benzocaine hydrochloride, isoflurane, sevoflurane, quinaldine sulfate, buffered tricaine methanesulfonate, 2-phenoxyethanol, injected pentobarbital, rapid chilling (appropriate zebrafish/research setting)	Eugenol, isoeugenol, clove oil, CO ₂ -saturated water (aquarium-fish facilities/fisheries), decapitation/cervical transection/manually applied blunt force trauma followed by pithing, rapid chilling followed by adjunctive method (aquarium-fish facilities), maceration (research setting)

Reptiles:

Acceptable	Acceptable With Conditions
As appropriate by species—Injected barbiturates, dissociative agents and anesthetics as specified	As appropriate by species—Inhaled anesthetics as specified, CO ₂ , penetrating captive bolt or firearm, manually applied blunt force trauma to the head, rapid freezing for animals < 4 g

Because it is often difficult to confirm that a reptile is dead, the application of two or more euthanasia procedures is usually recommended. Consulting multiple references on reptile euthanasia is advised as a means of identifying methods that are most appropriate for a given species and set of circumstances. (AVMA, 2013)

Nonhuman Primates:

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Acceptable	Acceptable With Conditions(as appropriate by species)
Intravenous barbiturates or anesthetic overdose	Inhaled anesthetics CO2 CO

Special Cases: When animals to be euthanized are fully anesthetized, adjunctive methods such as bilateral thoracotomy, exsanguination, perfusion, and IV or intracardiac injection of potassium chloride are acceptable.

Avian species:

Acceptable Methods	Acceptable with Conditions
Injected barbiturates Inhalant anesthetics CO ₂ CO Gunshot (free-ranging only)	N ₂ , Ar, Cervical dislocation Decapitation Thoracic compression (small free-ranging only) Maceration (chicks, and pipped eggs only]

Ruminants and Horses:

Acceptable Methods	Acceptable with Conditions
Injected barbiturates	Penetrating captive bolt* Gunshot to the head

Swine (Mature sows, Boars, and Grower-Finisher Pigs):

Acceptable Methods	Acceptable with Conditions
Injected barbiturates	CO ₂ CO N ₂ with CO ₂ Ar – alone or with CO ₂ Penetrating captive bolt* Gunshot to the head Electrocution

Swine (Nursery pigs – 70 lbs or lighter):

Acceptable Methods	Acceptable with Conditions
Injected barbiturates	CO ₂ (alone or in combination with N ₂ or Ar) CO Purpose-built nonpenetrating captive bolt

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	Electrocution (> 10 lb.)** Anesthetic overdose
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Swine (Suckling):

Acceptable Methods	Acceptable with Conditions
Injected barbiturates	CO2 (alone or with N2 or Ar) CO Inhaled anesthetics Purpose-built nonpenetrating captive bolt Electrocution (>10 lbs) Anesthetic overdose

Amphibians:

Acceptable Methods (as appropriate by species)	Acceptable with Conditions
Injected barbiturates Injected dissociative agents or anesthetics Topical buffered MS-222 Benzocaine hydrochloride	Inhaled anesthetics CO2 Penetrating captive bolt Blunt force trauma to the head Rapid freezing